



Record of piebaldism in *Dicotyles tajacu* (Artiodactyla: Tayassuidae) in northeast San Luis Potosí, Mexico

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Abstract

Phenotypic variations occur naturally in vertebrates. These variations are related to environmental, physiological, and genetic factors. Most variations in the pigmentation of the integument (skin, hair, feathers, scales, nails, etc.), or chromatic anomalies, originate due to the inheritance of an altered copy of genes related to the production of tyrosinase and melanin synthesis. Chromatic anomalies are classified as piebaldism, leucism, albinism, and melanism. This work presents the first record of piebaldism in the collared peccary (*Dicotyles tajacu*) in northeastern Mexico, being only the second record in the entire range. This finding was made during a monitoring conducted in 2023 at a site adjacent to the Sierra del Abra Tanchipa Biosphere Reserve, in northeastern San Luis Potosí, Mexico. This record contributes to the understanding of the distribution of chromatic anomalies in the Neotropical region, as well as their possible ecological implications.

Key words: chromatic anomalies, collared peccary, Neotropical region, piebaldism.

Resumen

Las variaciones fenotípicas ocurren de manera natural en los vertebrados. Estas variaciones se relacionan con factores ambientales, fisiológicos y genéticos. La mayoría de las variaciones en la pigmentación del integumento (piel, pelo, plumas, escamas, uñas, etc.), o anomalías cromáticas, se originan debido a la herencia de copias alteradas de genes relacionados con la producción de tirosinasa y síntesis de melanina. Las anomalías cromáticas se clasifican como piebaldismo, leucismo, albinismo y melanismo. En este trabajo se presenta el primer registro de piebaldismo en el pecarí de collar (*Dicotyles tajacu*) en el noreste de México, siendo apenas el segundo registro en toda su distribución. Este hallazgo se realizó durante un monitoreo realizado en 2023 en un sitio adyacente a la Reserva de la Biosfera Sierra del Abra Tanchipa, en el noreste de San Luis Potosí, México.

Este registro contribuye a la comprensión de la distribución de anomalías cromáticas en la región Neotropical, así como sus posibles implicaciones ecológicas.

Palabras clave: anomalías cromáticas, pecarí de collar, piebaldismo, región Neotropical.

Pigmentation variability represents one of the most prominent phenotypic characteristics observed in vertebrates (Hofreiter & Schöneberg 2010). Color patterning and variation result from the integrated function of more than 150 pigmentation-related genes. Sequence mutations within these genes drive phenotypic differences in pigmentation, contributing to visual attraction, aposematism, and camouflage (Hofreiter & Schöneberg 2010; Howell et al. 2021). The diversity of coloration patterns in mammals primarily arises from the presence and distribution of specific pigments—predominantly melanin—within the integumentary system (Pawelek & Körner 1982; Lucati & López-Baucells 2016; Fertl & Rosel 2018). These coloration patterns play key roles in adaptive processes, including sexual selection, intra- and interspecific communication, susceptibility to diseases, protection, predatory success, and anti-predator defense (Caro 2005; Caro & Mallarino 2020; Howell et al. 2021).

Nonetheless, chromatic anomalies can result from either an overproduction or deficiency of melanin, manifesting as partial or complete alterations in coloration across the body (Hofreiter & Schöneberg 2010; Abreu et al. 2013). Recent zoological studies have classified chromatic anomalies as piebaldism, leucism, albinism, and melanism (Acevedo et al. 2009; Lucati & López-Baucells 2016; Fertl & Rosel 2018). However, there is no absolute consensus because some authors use different terms or adjectives such as "partial" or "total" (Lucati & López-Baucells 2016; Zalapa et al. 2016), especially in cases of hypopigmentation (i.e. piebaldism, leucism, and albinism).

Piebaldism is defined as the absence of pigmentation only in specific parts of the body, while eye color remains normal (Acevedo et al. 2009; Lucati & López-Baucells 2016). It is an autosomal dominant disorder characterized by defective melanocyte migration into the affected skin and hair follicles or feathers due to mutations in the *KIT* gene, although pathogenetic variants in the *SNAI2* gene have also been described (Finch et al. 2017; Giacaman & Martín-Santiago 2023). Individuals with chromatic anomalies, such as piebaldism, may suffer numerous visual and immunological pathologies (Summers 2009) and would be more susceptible to predation (Sazima & Di-Bernardo 1991; Parsons & Bonderup-Nielsen 1995).

Even though chromatic anomalies have been reported in several Neotropical mammals (Abreu et al. 2013; Lucati & López-Baucells 2016; García et al. 2020; Marín 2021; Cotts et al. 2023, 2024; Gallo et al. 2023; Fuentes et al. 2024), these anomalies are considered rare in free-ranging populations due to the assumed short life expectancy of animals carrying these traits (Silva et al. 2020). In wild ungulates there are several records of chromatic anomalies, including cervids such as Amazonian brown brocket (*Passalites nemorivagus* Mejía-Valenzuela & Auz-Cerón 2020; Guastalla et al. 2021), gray brocket deer (*Subulo gouazoubira* Oliveira 2009; Nigro 2020; Guastalla et al. 2021; Aprile 2022; Muzzachiodi, 2023), marsh deer (*Blastocerus dichotomus* Pereira et al. 2023), red brocket deer (*Mazama americana* Mejía-Valenzuela & Auz-Cerón 2020; Ribeiro & Siqueira-Silva 2020; Guastalla et al. 2021), as well as lowland tapirs (*Tapirus terrestris* Alviz et al. 2005; Landis 2020; Tirira & Arbeláez 2020), collared (*Dicotyles tajacu* Veiga 1994; da Silva et al. 2019; Ruíz-Gutiérrez et

al. 2023; Leão-Vulcão et al. 2024; Urgilés-Verdugo et al. 2024) and white-lipped peccaries (*Tayassu pecari* Aximoff et al. 2021).

Collared peccaries are pig-like ungulates with a large, triangular head, a stocky, compact body, and small legs (total length: 93.2–103 cm; weight: 12–28 kg). These tayassuids have grayish-brown fur with a thin pale-yellow line from the shoulders to the neck (Reid 2006; Aranda 2012; Reyna-Hurtado et al. 2014). The species' distribution ranges from the southern United States (Arizona, New Mexico, and Texas) to Argentina. Collared peccaries are highly gregarious, forming groups ranging from 2–15 individuals in tropical deciduous forests to as many as 50 in desert areas (Reyna-Hurtado et al. 2014). Specifically, only one case of piebaldism has been reported in collared peccaries throughout their distribution; that record comes from Brazil (Leão-Vulcão et al. 2024). For Mexico, there is only one previous record in the scientific literature of chromatic anomalies—leucism—for this species (Ruíz-Gutiérrez et al. 2023), as well as a record of a pair of albino specimens documented on a citizen science platform (<https://mexico.inaturalist.org/observations/1122662>). Here, we report the second record of a free-ranging *D. tajacu* with piebaldism in the entire range.

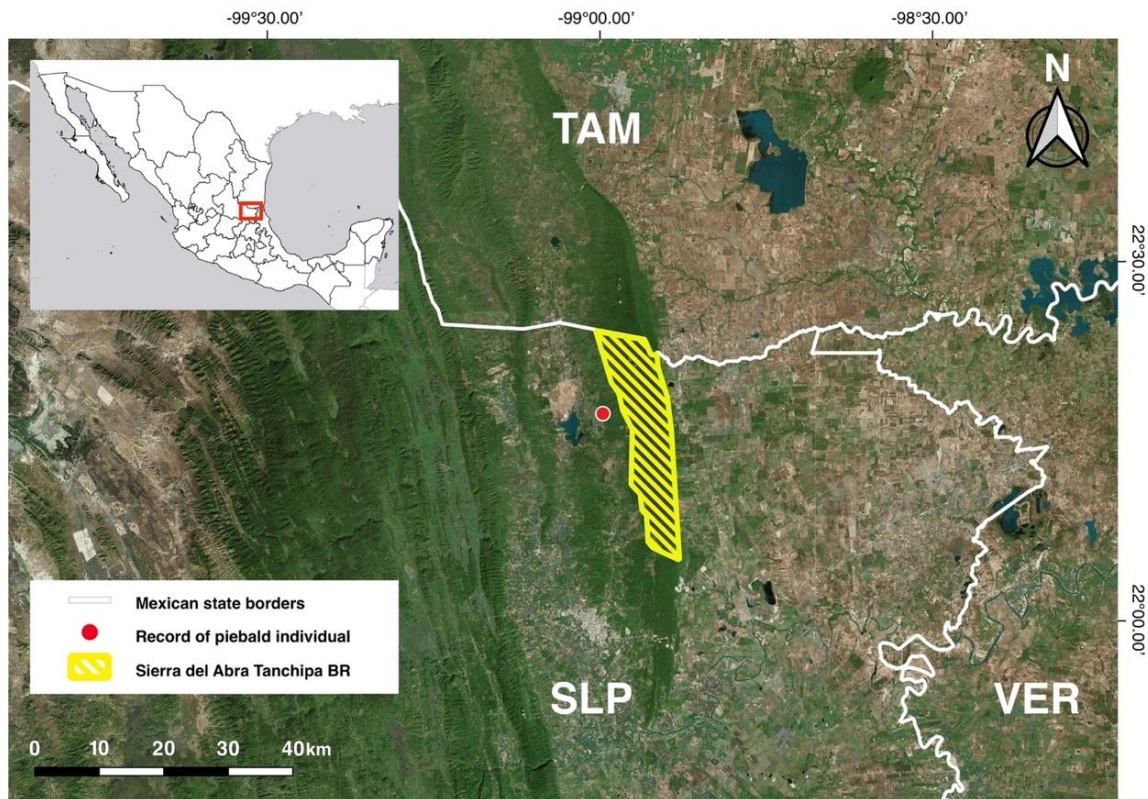


Figure 1. Location of the collared peccary (*Dicotyles tajacu*) individual with piebaldism (red dot) in the adjacent area of the Sierra del Abra-Tanchipa Biosphere Reserve (yellow striped polygon) in northeast San Luis Potosí, Mexico. Mexican states: TAM = Tamaulipas; SLP = San Luis Potosí; and VER = Veracruz.

The study site was the area adjacent to the Sierra del Abra-Tanchipa Biosphere Reserve (RBSAT), a natural protected area located in northeastern San Luis Potosí state (22,083–

22,4061, -98,8794–99,0167), with elevations ranging from 120 to 820 m (CONANP 2014). This area is considered a Neotropical stronghold in northeastern Mexico (Figure 1). The region receives an average annual rainfall of 1,070 mm, with 81% falling during the wet season (June–October). The mean annual temperature is 25.7°C, although temperatures can rise to 51°C during the dry season (November–May; Durán-Fernández et al. 2020). The main vegetation includes low and medium sub-perennial rainforest, tropical deciduous forest, tropical oaks (*Quercus oleoides*), palm (*Sabal mexicana*) groves, and secondary vegetation (CONANP 2014; Durán-Fernández et al. 2020). The RBSAT is bordered by agricultural valleys featuring sugarcane (*Saccharum* spp.), citrus, and sorghum (*Sorghum* spp.) plantations, as well as cattle pastures and human settlements (Chapa-Vargas & Monzalvo-Santos 2012).

The record was obtained through the opportunistic use of camera traps (Browning Trail Cam®–Prometheus Group LLC) as part of a nature photography project that documented the wildlife species that inhabit the region. The cameras were placed in a dry stream within the tropical deciduous forest and near several sugarcane fields (22,2886, -98,9956; WGS84); in the vicinity of the RBSAT (Figure 1). They were positioned approximately 50 cm above the ground and configured to take videos of 30-second length every twenty minutes. The cameras remained active 24 hours a day between May and November of 2023. During this period, we obtained a total effort of 1,225 camera days with 28 videos of a female *D. tajacu* with white body parts, totaling 19 independent records (24-hour interval). They were made between August 24 and November 5 and included only videos of the piebald animal with the group, with periods of daytime (06hr00–18hr00) and nighttime (19hr00–05hr00) activity. Non-pigmented parts of the specimen included the forehead and left side of the face, the back to shoulder height including the flanks and part of the forelimbs, as well as part of the rump (Figure 2).

Our results consistently show that, despite not having the normal greyish-brown coloration pattern (Reid 2006; Aranda 2012; Reyna-Hurtado et al. 2014), the female exhibiting piebaldism interacted normally with conspecifics. This interaction included olfactory exploration, scent-based contact with other individuals, and even suckling her offspring. Similar behavior has been observed in other tayassuids with chromatic anomalies (da Silva et al. 2019; Aximoff et al. 2021; Leão-Vulcão et al. 2024). This is because social cohesion in these species relies on vocal and olfactory signals (Byers & Bekoff 1981; Kiltie & Terborgh 1981; SOWLS 1997); therefore, individuals with different pigmentation would not be rejected from the herd (da Silva et al. 2019). Although they are considered to have a lower life expectancy (Sazima & Di-Bernardo 1991; Parsons & Bonderup-Nielsen 1995; Summers 2009), collared peccaries with chromatic anomalies that can reach adulthood have been registered (Veiga 1994; da Silva et al. 2019; Ruíz-Gutiérrez et al. 2023; Leão-Vulcão et al. 2024; Urgilés-Verdugo et al. 2024). Our record even documents that they can reproduce successfully.

Recently, chromatic anomalies have been frequently reported in mammal species, mainly those of medium or large size. However, little is known about the costs associated with these anomalies (Silva-Caballero et al. 2014), particularly hypopigmentation. Such anomalies are thought to provide indirect evidence of underlying genetic variability, as well as genetic changes related to fitness and population structure (Acevedo & Aguayo, 2008). Moreover, chromatic anomalies are frequently observed in gregarious or sedentary species and are thought to be more common in small, isolated populations (Bensch et al. 2000; Guevara et al. 2011). In recent decades, severe land-use change and habitat fragmentation have been documented in the RBSAT region (Painter et al. 2022), suggesting

a possible influence on the occurrence of this type of chromatic anomaly due to factors such as inbreeding (Bensch et al. 2000; Silva-Caballero et al. 2014), combined with the hunting pressure suffered by the species (Méndez et al. 2025).

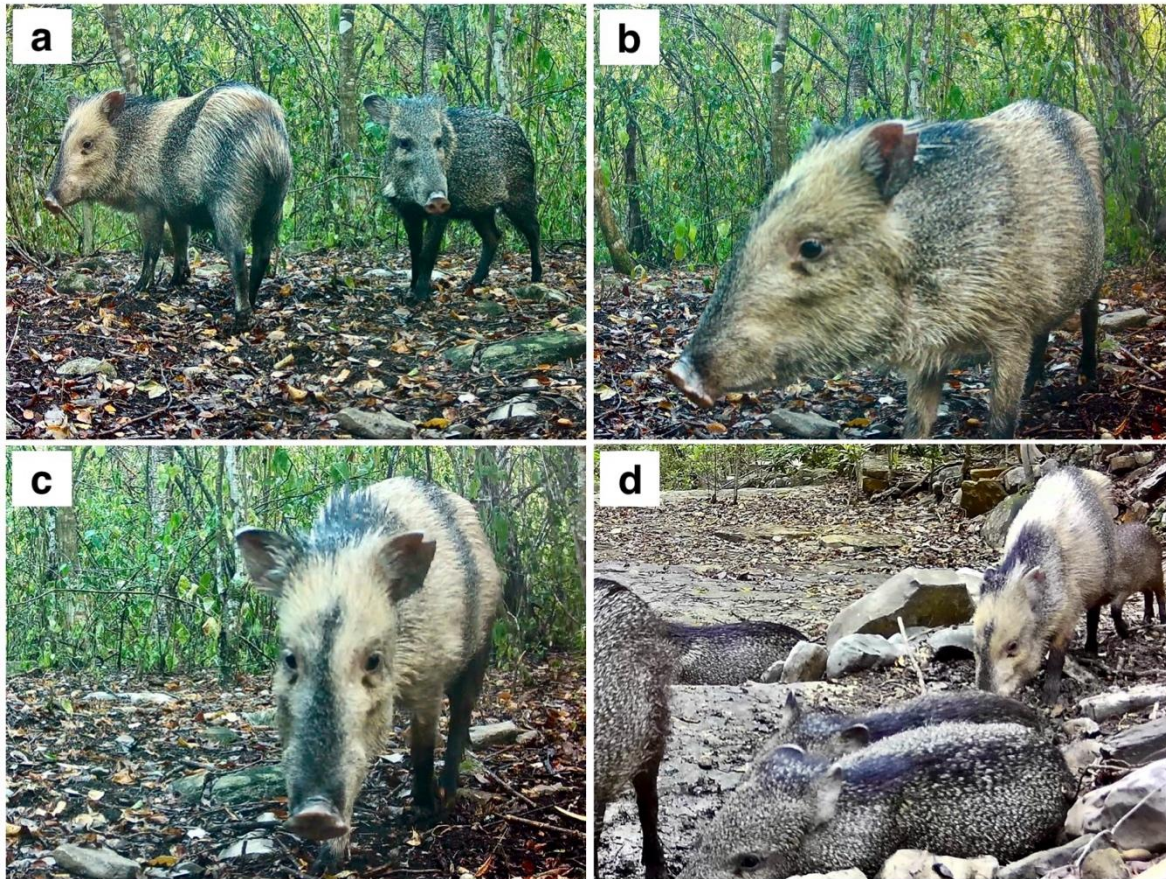


FIGURE 2. Female collared peccary (*Dicotyles tajacu*) with piebaldism: a) piebald individual (left) next to another individual with normal coloration, note the difference in fur coloration; b) and c) the non-pigmented parts of the specimen can be seen, including forehead and left side of the face, the back to shoulder height including the flanks and part of the forelimbs, as well as part of the rump; d) individual nursing its young.

Another possible explanation for the record presented here is that despite the low frequency of chromatic anomalies in free-ranging populations (Silva et al. 2020), they have been more commonly recorded in recent times due to greater access by more people to camera trapping tools (Palencia et al. 2022), which has allowed the recording of these variations in fur coloration, predation events, and behaviors never before recorded in wildlife populations. Therefore, we highlight the importance of camera traps as valuable tools for documenting the natural history of wildlife populations. These types of records should continue to be published to better understand this phenomenon, its distribution, and its potential ecological, physiological, and genetic implications.

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REFERENCES

- Abreu MSL, Machado R, Barbieri F, Freitas NS, Oliveira LR. 2013. Anomalous colour in Neotropical mammals: a review with new records for *Didelphis* sp. (Didelphidae, Didelphimorphia) and *Arctocephalus australis* (Otariidae, Carnivora). Brazilian Journal of Biology 73(1):185-194. <https://doi.org/10.1590/S1519-69842013000100020>
- Acevedo J, Aguayo M. 2008. Leucistic South American sea lion in Chile, with a review of anomalously color in otariids. Revista de Biología Marina y Oceanografía 43(2):413-417.
- Acevedo J, Torres D, Aguayo-Lobo A. 2009. Rare piebald and partially leucistic Antarctic fur seals, *Arctocephalus gazella*, at Cape Shirre, Livingston Island, Antarctica. Polar Biology 32(1):41-45. <http://doi.org/10.1007/s00300-008-0500-6>
- Alviz A, Quintero G, Farfán-Ardila N, Pérez-Albarracín K. 2025. First record of leucism in a wild lowland tapir *Tapirus terrestris* (Linnaeus, 1758) from the Orinoquia region of Colombia. Notas sobre Mamíferos Sudamericanos 7(1):2-8. <http://doi.org/10.31687/SaremNMS25.1156>
- Aprile G. 2022. Primer reporte documentado de piebaldismo en *Mazama gouazoubira* (Artiodactyla: Cervidae) en la provincia de Córdoba, Argentina. Acta Zoológica Lilloana 66(2):197-204. <https://doi.org/10.30550/j.azl/2022.66.2/2022-09-19>
- Aranda, M. 2012. Manual para el rastreo de mamíferos silvestres de México. Distrito Federal, México: Comisión Nacional para el Conocimiento y Uso de la Biodiversidad.
- Aximoff I, Neto EP, de Paula W, Hofmann GS, Keuroghlian A, Jorge ML, Lima E, Barquero G. 2021. Sticking out in a herd? Records of anomalous pigmentation in a social herd-forming ungulate (*Tayassu pecari*). North-Western Journal of Zoology 17(2):288-293.
- Bensh S, Hansson B, Hasselquist D, Nielsen B. 2000. Partial albinism in a semi-isolated population of great reed warblers. Hereditas 133(2):167-170. <https://doi.org/10.1111/j.1601-5223.2000.t01-1-00167.x>
- Byers JA, Bekoff M. 1981. Social, spacing, and cooperative behavior of the collared peccary, *Tayassu tajacu*. Journal of Mammalogy 62(4):767-785. <https://doi.org/10.2307/1380598>
- Caro T. 2005. The adaptive significance of coloration in mammals. BioScience 55(2):125-136. [https://doi.org/10.1641/0006-3568\(2005\)055\[0125:TASOCI\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2005)055[0125:TASOCI]2.0.CO;2)
- Caro T, Mallarino R. 2020. Coloration in mammals. Trends in Ecology & Evolution 35(4):357-366. <https://doi.org/10.1016/j.tree.2019.12.008>
- Chapa-Vargas L, Monzalvo-Santos K. 2012. Natural protected areas of San Luis Potosi, Mexico: ecological representativeness, risks, and conservation implication across scales. International Journal of Geographical Information Science 26(9):1625-1641. <https://doi.org/10.1080/13658816.2011.643801>
- CONANP (Comisión Nacional de Áreas Naturales Protegidas). 2014. Programa de Manejo Reserva de la Biosfera Sierra del Abra Tanchipa. Ciudad de México, México: Secretaría de Medio Ambiente y Recursos Naturales.

- Cotts L, Slifkin JP, Moratelli R, Gonçalves L, Rocha-Barbosa O. 2023. Multiple colors in anteaters: review and description of chromatic disorders in *Tamandua* (Xenarthra: Pilosa) with reports of new and rare coat colorations. *Zoologia* (Curitiba) 40:e22034. <https://doi.org/10.1590/S1984-4689.v40.e22034>
- Cotts L, Costa G, Gabriel S, Rocha-Barbosa O, Braga C. 2024. Unveiling the colors of mustelids: a historical review on the emergence of chromatic disorders and their ecological effects in mustelids worldwide with report of the first case of erythrism in *Eira barbara* (Carnivora, Mustelidae). *Animals* 14(23):3354. <https://doi.org/10.3390/ani14233354>
- da Silva VL, Cândido J, Campanha JN, de Oliveira DR, Gheler-Costa C, Oda FH. 2019. A stranger in the family? On the social behavior of a leucistic collared peccary (*Pecari tajacu*) with pigmented conspecifics. *Tropical Ecology* 60:303–305. <https://doi.org/10.1007/s42965-019-00036-x>
- Durán-Fernández A, Silva-Caballero A, Montoya-Gandarillas S, Godínez-Vizuet O. 2020. Historia y conservación del jaguar en la Sierra del Abra Tanchipa. In: Rosas-Rosas OC, Silva-Caballero A, Durán-Fernández A, editors. Manejo y conservación del jaguar en la Reserva de la Biosfera Sierra del Abra Tanchipa. Texcoco, México: Colegio de Postgraduados. p. 61–85.
- Fertl D, Rosel P. 2018. Albinism. In: Würsig B, Thewissen JGM, Kovacs KM, editors. *Encyclopedia of Marine Mammals*. 3rd ed. New York: Academic Press. p. 20–21.
- Finch J, Abrahams S, Finch A. 2017. Analogs of human genetic skin disease in domesticated animals. *International Journal of Women's Dermatology* 3(3):170–175. <https://doi.org/10.1016/j.ijwd.2017.01.003>
- Fuentes R, Castillo M, Moreno R, Quintero-Arrieta H, Pérez E, Araúz J, Añino Y, Murcia-Moreno D, Valdés R, Bonilla B, Gálvez D. 2024. Report of coloration anomalies in mammals from Panama. *Neotropical Biology and Conservation* 19(1):333–345. <https://doi.org/10.3897/neotropical.19.e125890>
- Gallo JA, Pagnutti N, Chiale MC, Abba AM. 2023. Anormalidades cromáticas en armadillos (Xenarthra:Cingulata): registros hallados en la web. *Notas sobre Mamíferos Sudamericanos*. 5(1):2–9. <https://doi.org/10.31687/saremnms23.8.4>
- García FJ, Machado M, Flórez G, Soriano PJ, Cabrer HJ. 2020. Anomalous coloration in Neotropical mammals: New cases of pigmentation disorders in Venezuelan mammals revealed by review of natural history museums. *Memoria de la Fundación La Salle de Ciencias Naturales* 78(186):5–20.
- Giacaman A, Martín-Santiago A. 2025. Piebaldismo. *Actas Dermo-Sifiliográficas* <https://doi.org/10.1016/j.ad.2023.12.009>
- Guastalla MG, Azevedo FC, Vogliotti A, Lacerda WR, Magina GC, Cardoso CAC, Endo W, Peres CA, Haugaasen T, Rocha DG, Lemos FG. 2021. The mysterious white deer: anomalous coloring in different Neotropical deer. *Mammalian Biology* 101:665–673. <https://doi.org/10.1007/s42991-021-00156-9>
- Guevara L, Ramírez-Chaves HE, Cervantes FA. 2011. Leucismo en la musaraña de orejas cortas *Cryptotis mexicana* (Mammalia: Soricomorpha), endémica de México. *Revista Mexicana de Biodiversidad* 82(2):731–733. <https://doi.org/10.22201/ib.20078706e.2011.2.1194> guevara
- Hofreiter M, Schöneberg T. 2010. The genetic and evolutionary basis of colour variation in vertebrates. *Cellular and Molecular Life Science* 67:2591–2603. <https://doi.org/10.1007/s00018-010-0333-7>
- Howell N, Sheard C, Koneru M, Brockelsby K, Ono K, Caro T. 2021. Aposematism in mammals. *Evolution* 75(10):2480–2493. <https://doi.org/10.1111/evo.14320>

- Kiltie RA, Terborgh J. 1983. Observations on the behavior of rain forest peccaries in Perú: Why do white-lipped peccaries form herds?. *Zeitschrift für Tierpsychologie* 62(3):241-255.
- Landis MB, Candisani L, Munhoes LP, Gebin JCZ, Rezende F, Flores de Jesus, MM, Medici EP, Ferraz KMPMB. 2020. First record of albino lowland tapirs (*Tapirus terrestris* Linnaeus 1758) in an important Brazilian Atlantic Forest hotspot. *Mammalia* 84(6):601-604. <https://doi.org/10.1515/mammalia-2019-0084>
- Leão-Vulcão OOP, do Val HGP, Brasil GR, Pezzuti JCB, Carvalho Jr EAR. 2024. First record of White-Collared Peccary (*Dicotyles tajacu*) with piebaldism across its entire distribution. *Mammalogy Notes* 10(2):450. <https://doi.org/10.47603/mano.v10n2.450>
- Lucati F, López-Baucells A. 2016. Chromatic disorders in bats: a review of pigmentation anomalies and the misuse of terms to describe them. *Mammal Review* 47(2):112-123. <https://doi.org/10.1111/mam.12083>
- Marín, D. 2021. Coloración atípica del pelaje en algunos pequeños mamíferos no voladores de Colombia. *Mammalogy Notes* 7(1):180. <https://doi.org/10.47603/mano.v7n1.180>
- Mejía-Valenzuela EG, Auz-Cerón DA. 2020. Primer reporte de leucismo en Mazama murelia y Mazama zamora (Artiodactyla: Cervidae) en el Corredor Cuyabeno-Yasuní, Ecuador. *Notas sobre Mamíferos Sudamericanos* 2(1):2-7. <http://doi.org/10.31687/saremNMS.20.0.05>
- Méndez MAB, Pérez RM, Cordero WA, Cruz AP. 2011. Cacería del pecarí de collar (*Pecarí tajacu*) (Artiodactyla: Tayassuidae) en tzucacab, Yucatán, México. *Revista Mexicana de Biodiversidad* 1(1):8-18. <https://doi.org/10.22201/ie.20074484e.2011.1.1.9>
- Muzzachiodi N. 2023. Leucismo parcial en cría de *Subulo gouazoubira* (Mammalia:Artiodactyla:Cervidae) en el Norte entrerriano. *Acta Zoológica Platense* 9(85):1-4.
- Nigro NA, Lodeiro Ocampo N, Falke F, Braslavsky OH. 2020. Hallazgo de tres ejemplares de corzuela parda (*Mazama gouazoubira*) con leucismo parcial en el Parque Nacional Baritú, Provincia de Salta, Argentina. *Nótulas Faunísticas (Segunda Serie)* 287:1-7.
- Oliveira SF. 2009. Registro de albinismo parcial em veado catingueiro *Mazama gouazoubira* (G. Fischer, 1814) (Artiodactyla, Cervidae) na Serra do Sudeste, Rio Grande do Sul, Brasil. *Biodiversidad Pampeana* 7(1):13-15.
- Parsons GJ, Bonderup-Nielsen S. 1995. Partial albinism in an island population of Meadow Voles, *Microtus pennsylvanicus*, from Nova Scotia. *The Canadian Field Naturalist* 109(2):263-264. <https://doi.org/10.5962/p.357626>
- Painter EJ, Rosas-Rosas OC, Bender LC, Tarango-Arámbula LA, Martínez-Montoya JF, Guerrero-Rodríguez JD, Silva-Caballero A. 2022. Land use change and its implications for biodiversity and jaguar conservation. *Therya* 13(3):277-286. <https://doi.org/10.12933/therya-22-2114>
- Palencia P, Vicente J, Soriguer RC, Acevedo P. 2022. Towards a best-practices guide for camera trapping: assessing differences among camera trap models and settings under field conditions. *Journal of Zoology* 316(3):197-208. <https://doi.org/10.1111/jzo.12945>
- Pawelek J.M, Körner A.M. 1982. The Biosynthesis of Mammalian Melanin: The regulation of pigment formation, the key to disorders such as albinism and piebaldism, may also offer some clues for the treatment of melanoma. *American Scientist* 70(2):136-145.
- Pereira JA, Wolfenson LI, Artero DF, Argerich EC, Varela D, Fracassi NG. 2023. First records of pigmentation anomalies in the marsh deer at its southernmost stronghold: a warning message?. *European Journal of Wildlife Research* 69:59. <https://doi.org/10.1007/s10344-023-01694-y>

- Reid, F. 2006. A field guide to mammals of North America. New York, USA: Houghton Mifflin Harcourt.
- Reyna-Hurtado R, March I, Naranjo E, Mandujano S. 2014. Pecaríes en México. In: Valdéz R, Ortega-S A ,editors. Ecología y manejo de fauna silvestre en México. Texcoco, México: Colegio de Postgraduados. p. 353-375.
- Ribeiro R, Siqueira-Silva DH. 2020. First report of complete albinism in *Mazama americana* (Erxleben, 1777) in the Biological Reserve of Tapirapé, Oriental Amazon, Brazil. *Acta Scientiarum (Biological Sciences)* 42:e46734.
- Ruíz-Gutiérrez F, Chávez C, Hernández-Hernández J, Torres-Bernal R, Ruiz-Gutiérrez B, Vázquez-Arroyo E, Barrera-Nava E, Samudio-Pineda O. 2023. Reporte de *Pecari tajacu* y *Procyon lotor* con anomalías cromáticas en el Estado de Guerrero, México. *Revista Peruana de Biología* 30(2):e24148. <http://doi.org/10.15381/rpb.v30i2.24148>
- Sazima I, Di-Bernardo M. 1991. Albinismo em serpentes neotropicais. *Memórias do Instituto Butantan* 53(2):167–173.
- Silva FA, Lessa G, Bertuol F, Freitas TRO, Quintela FM. 2020. Chromatic anomalies in Akodontini (Cricetidae: Sigmodontinae). *Brazilian Journal of Biology* 80(2):479–481. <https://doi.org/10.1590/1519-6984.214680>
- Silva-Caballero A, Montiel-Reyes F, Sánchez-Garibay E, Ortega J. 2014. Leucismo en el coatí de nariz blanca *Nasua narica* (Mammalia: Carnivora), en Quintana Roo, México. *Therya* 5(3):839-843. <https://doi.org/10.12933/therya-14-193>
- Sowls LK. 1997. Javelinas and the other peccaries: their biology management and use. 2nd ed. College Station, Texas: Texas A&M University.
- Summers CG. 2009. Albinism: classification, clinical characteristics, and recent findings. *Optometry and Vision Science* 86(6):659–662. <https://doi.org/10.1097/OPX.0b013e3181a5254c>
- Tirira DG, Arbeláez E. 2020. Primer reporte de leucismo en un tapir amazónico (Perissodactyla, Tapiridae) nacido bajo cuidado humano. *Mammalia aequatorialis* 2020(2):85-88. <https://doi.org/10.59763/mam.aeq.v2i.19>
- Urgilés-Verdugo C, Payaguaje L, Biaguaje A, Payaguaje H, Esbach MS. 2024. Anomalías en la coloración de piel y pelo de *Dicotyles tajacu* (Artiodactyla: Tayassuidae) y *Dasyprocta fuliginosa* (Rodentia: Dasyproctidae) en la Amazonía ecuatoriana. *Mammalogy Notes*, 10(2):412. <https://doi.org/10.47603/mano.v10n2.412>
- Veiga LA. 1994. Um caso de albinismo em *Tayassu tajacu* Linnaeus (Artiodactyla, Tayassuidae) na Serra do Mar, São José dos Pinhais, Paraná. *Revista Brasileira de Zoologia* 11: 341-343. <https://doi.org/10.1590/S0101-81751994000200019>
- Zalapa SS, Guerrero S, Romero-Almaraz ML, Sánchez-Hernández C. 2016. Coloración atípica en murciélagos: frecuencia y fenotipos en Norte y Centroamérica e islas del Caribe y nuevos casos para México y Costa Rica. *Revista Mexicana de Biodiversidad* 87(2):474-482. <https://doi.org/10.1016/j.rmb.2016.04.007>

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